

## **Analgesic And Neuropharmacological Evaluation of Ethyl Acetate Extract from *Kalanchoepinnata* leaves**



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**Declaration by the Research candidate**

I, Md. JamilUddin, hereby declare that the dissertation entitled “**Neuropharmacological Evaluation of Ethyl Acetate Extract from *Kalanchoepinnata* leaves**”, submitted by me to the Department ofPharmacy,East West University, in partial fulfillment of the requirements for the award of the degree of Bachelor of Pharmacy (B.PHARM) is a complete record of original research work carried out by me during the period 2011-2012 under the supervision and guidance of Md. RazibulHabib, Lecturer, Department of Pharmacy, East West University and it has not formed the basis for the award of any other Degree/Diploma/Fellowship or other similar title to any candidate of any University.

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## Thesis Certificate

This is to certify that the thesis entitled “**Neuropharmacological Evaluation of ethyl Acetate Extract from *Kalanchoepinnata*leaves**”, submitted to the Department of Pharmacy, East West University, in partial fulfillment of the requirements for the award of the degree of Bachelor of Pharmacy (B.PHARM) is a complete record of original research work carried out by Md. JamilUddin (ID. 2007-3-70-045) during the period 2011-2012 of his research in the Department of Pharmacy at East West University, under my supervision and guidance and the thesis has not formed the basis for the award of any other Degree/Diploma/Fellowship or other similar title to any candidate of any University.

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**ABSTRACT**

The analgesic and Neuropharmacological activity of acetic acid extract of leaves of *Kalanchoepinnata*(Lam.) Pers. was evaluated using an acetic acid-induced gastric pain model in Swiss albino mice. The acetic acid extract of leaves of *Kalanchoepinnata* demonstrated significant dose-dependent analgesic activity at all the tested doses of 100, 200, and 400 mg leaf extract/kg body weight in mice. Even at the lowest dose of 100 mg/kg body weight, the analgesic activity of leaf extract was comparable to that of a standard analgesic drug, aspirin, administered at 200 mg/kg body weight. The highest inhibition of writhing induced by acetic acid (47.5%) was observed with a dose of 400 mg leaf extract/kg body weight, which was much greater than the inhibition obtained with aspirin (38.4%). At the lower dose of 250 mg bark extract/kg body weight, there was a decrease in the number of writhings compared to controls, but the decrease was not significant. The results suggest that acetic acid extract of leaves of *Kalanchoepinnata* and chloroform extract of barks have activities and validates of folk medicinal uses in Bangladesh for treatment of pain. It was however more efficacious against picrotoxin-induced seizure where protection was observed in about one-quarter of mice, an effect which indicates that *k. pinnata* aqueous extract might produce its central nervous system depressant action as consequence of its GABAergic and less importantly, glycinergic transmission, since picrotoxin is a selective GABAA receptor antagonist (Rang et al 1996) while strychnine antagonizes the inhibitory spinal cord and brainstem reflexes of glycine (Yemitan et al 2001).

Key words: *Kalanchoepinnata*, Analgesic test, Writhing test, Formalin induced test, Neuropharmacology, Hole cross, Open field, EPM test, Result

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## Introduction

*Kalanchoepinnata*(Lam.) Pers. (Family: Crassulaceae, local name: Pathorkuchi) is an herb foundubiquitously in Bangladesh having thick succulent leaves. Leaves have a variety of uses in the traditionalmedicinal system of Bangladesh. They are eaten for diabetes control, used as a diuretic and for dissolving ofkidney stones, and taken for respiratory tract infections, as well as applied to wounds, boils, and insect bites(Ghani, 2003). The leaves are used by traditionalmedicinalpractitioners for diabetes, and the bark as a purgative (Ghani, 2003).Phenolic components (Gaind and Gupta, 1973) and flavonoid glycosides (Gaind and Gupta, 1971) havebeen reported from *Kalanchoepinnata*. The leaf extract of the plant has been shown to significantly delay theonset of leishmaniasis in BALB/c mice when infected with *Leishmaniaamazonensis*(Da Silva *et al.*, 1995). Quercitrin (quercetin 3-O-a-L-rhamnopyranoside), obtained from an active aqueous extract of the plant has been identified as the active principle (Muzitano*et al.*, 2006). Effectiveness of an immunomodulatory extract of the plant has also been demonstrated against visceral leishmaniasis in BALB/c mouse model of infection with*Leishmaniachagasi*(Gomes *et al.*, 2010). Other activities reported for the plant or plant components include protection of mice against fatal anaphylactic shock (Cruz *et al.*, 2008), hepatoprotective activity in leaves (Yadav and Dixit, 2003), anti-tumor promoting activity of bufadienolides isolated from leaves of the plant (Supratman*et al.*, 2001) and isolation of insecticidal bufadienolides from leaves of the plant (Supratman*et al.*, 2000). The anti-diabetic and anti-obesity activity of *Lagerstroemia speciosa*as well as several phytochemical constituents isolated from the plant has been very well documented (Kakuda*et al.*, 1996; Suzuki *et al.*, 1999;Liu *et al.*, 2001; Hayashi *et al.*, 2002; Judy *et al.*, 2003; Hattori *et al.*, 2003; Liu *et al.*, 2005; Yamada *et al.*, 2008; Klein *et al.*, 2007; Bai*et al.*, 2008; Houet *et al.*, 2009; Sivakumare*et al.*, 2009; Ichikawa *et al.*, 2010).A phytochemical isolated from the plant, orobol 7-O-D-glucoside, has been reported to have inhibitory effectson human rhinoviruses replication (Choi *et al.*, 2010). Free radical scavenging and anti-inflammatory propertieshave been demonstrated in leaf extracts of the plant (Priya*et al.*, 2008). Ethyl acetate extract of leaves hasbeen shown to ameliorate cisplatin-induced nephrotoxicity in BALB/c mice (Priya*et al.*, 2007). Anti-fungal activity has been demonstrated with hot water as well as methanol extract of the plant against *Arthriniumsacchari*M001 and *Chaetomiumfunicola*M002 strains (Sato *et al.*, 2000). Xanthine oxidase inhibitors(valoneic acid dilactone and ellagic acid) have been isolated from leaves of the

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plant (Unnoet al., 2004).Petroleum ether extract of seeds reportedly showed antibacterial activity against both Gram positive and Gramnegative bacteria (Sinhababuet al., 1994).

In our ongoing survey on folk medicinal practices in Bangladesh (Rahmatullahet al., 2009a; 2009b; 2009c), it was observed that leaves of *Kalanchoepinnata*were used by the folk medicinal practitioners in several parts of Bangladesh for treatment of rheumatoid arthritis and pain in the bones (unpublished observation). The barks of *Lagerstroemia speciosa*were also used as an analgesic (unpublished observation). The objective of the present study was to determine the analgesic potential of leaves of *Kalanchoepinnata* whether the experimental results validate the plants' folk medicinal uses.

Oxygen radicals, the products of some biochemical and physiological reactions, initiate cell signaling pathways, damage cellular lipids, proteins, and nucleic acids. Reactive oxygen species are pivotal for the onset of various conditions such as hypertension, atherosclerosis, cancer, and alzheimer's disease (Morris, 2005; Azadzoiet al., 2005). During normal aerobic metabolism, activated oxygen radicals, hydroxyl radicals and peroxy nitrite. The *Kalanchoe*genus is a succulent perennial plant belonging to the class *Bryophyllum*. It grows naturally throughout the temperate areas of the world. In Africa, over 200 species have being identified and many of thesespecies have been used medicinally especially *Kalanchoepinnata*and *Kalanchoeintegra*(Figure 1A andB) which have been used traditionally for the treatment of many disease conditions like peptic ulcer, upper respiratory tract infections, coughs and as anti-infective in Ghana (Dokosi, 1998 and Torres-Santos et al., 2003 ). They grow widely along footpaths and forests in Ghanaand is called by various local names (for example, the Ewes call it 'aflatoga', Fantes -'eporow', Twi and Ga - 'egoro' and 'tamiwu' respectively). *Kalanchoe*is reported to contain considerable amounts of flavonoid and phenolic compounds (Gaind and Gupta, 1971; Adenike and Eretan, 2004). Despite its rich flavonoid and phenolic content, available literature indicates that no study has been carried out to investigate the antioxidant properties of this medicinal plant. The aim of this study is to determine the total flavonoid and phenolic content of two *Kalanchoe* species, namely, *K. pinnata*and *K. integra*and also to assess their antioxidant activity.





**Fig: Pathorkuchi leaves (*Kalanchoepinnata*)**

Traditional medicine involves the use of herbal medicine, animal parts and minerals. However, herbal medicines are the most widely used of the three. Herbal medicines contain an active ingredient, aerial or underground parts of plants as their petal or seeds materials or combinations thereof, whether in the crude state or as plant preparations. Furthermore, about 80% of the world population is dependent (wholly or partially) on plant-based drugs (WHO, 1996). In Nigeria and most developing countries of the world, rural and urban dwellers, literate or illiterate rely heavily on herbal preparations for the treatment of various diseases despite the availability of orthodox medicine (Nwabuisi, 2002). *Bryophyllumpinnatum*(Lam.) (synonym: *Kalanchoepinnata*, Lam.; common names: Life plant, air plant (Mexican), love plant, Canterbury bells, Cathedral bells, e.t.c) is a perennial herb growing widely and used in folkloric medicine in tropical Africa, India, China, Australia and tropical America (Engler, 1926; Balzer et al, 1949). Classified as a weed(Oliver-Bever, 1983), the plant flourishes throughout the Southern part of Nigeria (Gill, 1992). A number of active compounds, including flavonoids, glycosides, steroids, bufadienolides and organic acids, have been identified in *Bryophyllumpinnatum*(Marriage and Wilson, 1971; Gaiind and Gupta, 1972; 1974; Costa et al, 1995). In traditional medicine, the leaves of this plant have been reported to possess antimicrobial (Mehta and Bhat, 1952; Akinpelu, 2000; Oliver-Bever, 1983), antifungal (Misra and Dixit, 1979), anti-ulcer (Pal and Nag, 1991), anti-inflammatory and analgesic (Pal and Nag, 1989; 1992), and antihypertensive (Ojewole 2002) activities. The methanol extract of the leaf of the plant has also been reported to have histamine receptor (H1) antagonism in the ileum, peripheral vasculature and bronchial muscle (Pal et al, 1999)

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Although, studies have shown the relative important effect of some medicinal plant on the central nervous system activities. (Dorr et al., 1971; Fujimori, 1995; Wakeel, 2004).. *B. pinnatum* has been used since 1921 in traditional medicine as an antipsychotic agent. Furthermore, Pal and Nag (1999) provided evidence for the neuropsychopharmacologic activities of the plant. The present study is therefore designed to further investigate the effects of the aqueous leaf extract of *B. pinnatum* on some central nervous. *Kalanchoepinnata* (Lamarck) Persoon (= *Bryophyllum pinnatum*) is a perennial medicinal herb, popularly used in Brazil and other parts of the world to treat various inflammatory diseases (Rossi-Bergmann et al., 1994).

Previous studies on the chemical composition of *K. pinnata* showed that bufadienolides, terpenoids and flavonoids are the main secondary metabolites of this species (Yamagishi et al., 1989; Costa et al., 1995). Our interest in *K. pinnata* is justified by its significant immunosuppressive effects, as well as its ability to protect against progressive infection with *Leishmania amazonensis* (Rossi-Bergmann et al., 1994; Da Silva et al., 1995). The leishmaniasis are a complex of diseases caused by different species of the protozoan parasite *Leishmania* and are a major public health problem in many developing countries, where 350 million people live at risk of infection (WHO, 2005). There is no approved vaccine for clinical use. Despite a few research achievements, first-line chemotherapy is still based on pentavalent antimonials, developed more than 50 years ago, which are toxic and prone to drug resistance (Croft and Coombs, 2003). Recently, several natural products with antileishmanial activity, including naphthoquinones, lignans, neolignans, alkaloids and triterpenoids have been reported (Chan-Bacab and Peña-Rodríguez, 2001).

However, there have been few studies on the antileishmanial activity of the flavonoid class of natural polyphenols. These few studies include that of luteolin, a common flavonoid in the human diet, which was recently described as a promising antileishmanial drug (Mittra et al., 2000). Proanthocyanidins also show antileishmanial activity, as well as modulatory effects on nitric oxide and tumor necrosis factor  $\alpha$  release in RAW 264.7 cells (Kolodziej et al., 2001), and a methoxychalcone isolated from inflorescences of *Piper aduncum* (Piperaceae) reportedly has significant antileishmanial activity as well (Torres-Santos et al., 1999). Quercitrin, previously isolated from an active flavonoid fraction of *K. pinnata* by our group, was an additional potent antileishmanial compound, with a low toxicity profile (Muzitano et al., 2006). Herein, we

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describe a new flavonoid diglycoside and two other polar flavonoids from *K. pinnata* as well as their in vitro antileishmanial activity in comparison with three analogs: quercetin, quercitrin, and afzelin.

## 1.1 Plant description of *kalanchoepinnata*

**Botanical Name:** *Bryophyllum pinnatum*

**Family Name:** Crussulaceae

**Sanskrit Name:** Pashanabheda

**Hindi Name:** Patharchur

**Common Names:** Cathedral Bells, Air Plant (USA), Life Plant, Miracle Leaf, Goethe Plant and Katakataka. Also called “Wonder of the World” in the English speaking Caribbean. 'OliwaKaKahakai (Hawai'i), Mother Of Thousands, Herbe Mal Tete (Dominica) Never Dead, Parvu, HojaDelAire (Bolivia).

**Synonym:** *Bryophyllum calycinum*, *Bryophyllum pinnatum*<sup>4,5,6</sup>.

## 1.2 Taxonomical tree

**Kingdom:** Plantae

**Division:** Magnoliophyta

**Class:** Magnoliopsida

**Order:** Saxifragales

**Genus:** *Kalanchoe*

**Section:** *Bryophyllum*

**Species:** *K. pinnata*

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The plant grows all over India in hot and moist areas, especially in Bengal. It is a succulent perennial plant that grows 1-1.5 m in height and the stem is hollow four-angled and usually branched. Leaves are opposite, decussate, succulent, 10-20 cm long. The lower leaves are simple, whereas, the upper ones 3-7 foliate and are long-petioled. They are fleshy dark green that are distinctively scalloped and trimmed in red. Leaf blade pinnately compound with 3-5 leaflets, 10-30 cm; petiolules 2-4 cm; leaflet blades oblong to elliptic, 6-8 X 3-5 cm, margin crenate with each notch bearing a dormant bud competent to develop into a healthy plantlet apex obtuse<sup>8</sup>. The leaves are furnished with rooting vegetative buds. Inflorescences terminal paniculate 10-40 cm. Flowers are many bell-like pendulous. Calyx tubular, 2-4 cm; Corolla reddish to purple, 5 cm, base sparsely ciliate; lobes ovate-lanceolate; stamens inserted basally on corolla; nectar scales oblong; follicles included in calyx and corolla tube. The fruit-pod with four septa and numerous, ellipsoid, smooth striate seeds within. The plant flowers in Nov-Mar and fruits in April 7,8,9 . It is astringent, sour in taste, sweet in the post digestive effect and has hot potency.



Figure: various plant parts of *k.pinnata*

**Habitat:** It is a succulent plant native to Madagascar. It is distinctive for the profusion of miniature plantlets that form on the margins of its leaves, a trait it has in common with the other members of the Bryophyllum section of the Kalanchoe genus. It is a popular houseplant and has become naturalized in temperate regions of Asia, the Pacific and Caribbean<sup>4</sup>.

**Distribution:** *Kalanchoe pinnata* has become naturalized in temperate regions of Asia, Australia, New Zealand, West Indies, Macaronesia, Mascarenes, Galapagos, Melanesia, Polynesia, and Hawaii. In many of these, such as Hawaii, it is regarded as an invasive species. It is also widely distributed in the Philippines and it is known as *katakataka* or *kataka-taka* which is also an adjective meaning astonishing or remarkable<sup>4,5</sup>.

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**Herbs as antibiotic:** The development of drug resistance in human pathogens against commonly used antibiotics has necessitated a search for new antimicrobial substances from other sources, including plants. Natural substances of plant origin have been used and are being used throughout the world for human and animal health care. The limitation of the synthetic medicines has created a challenge for the search of natural source of cure for many diseases. Plants used for traditional medicine contain a wide range of substances that are used to treat chronic as well as infectious diseases.

Herbal treatments are very effective in treating bacterial and other common childhood infections. Several herbs such as garlic, goldenseal, myrrh, usnea, and uvaursi have antibiotic effects. The difference between an antibiotic drug and an antibiotic herb is that the drug is an isolated constituent limited to the power of that one chemical, whereas the herb contains several constituents with a variety of healing properties, producing a synergistic effect.

The herb can actually kill only the bad bacteria while not harming the good; the drug does not have the wisdom to differentiate. Furthermore, most bacteria are not fooled by all isolated compound; Often the drug becomes ineffective or the cells mutate eventually to become resistant to the drug. The organic herb is nature's match for the bacteria.

### **Therapeutic Benefit**

Much of the exploration and utilization of natural products as antimicrobials arise from microbial sources. It was the discovery of penicillin that led to later discoveries of antibiotics such as streptomycin, aureomycin and chloromycetin. Though most of the clinically used antibiotics are produced by soil microorganisms or fungi, higher plants have also been a source of antibiotics. Examples of these are the bacteriostatic and antifungicidal properties of Lichens, the antibiotic action of allinine in *Allium sativum* (garlic), or the antimicrobial action berberines in goldenseal (*Hydrastiscanadensis*)

### **Economic Benefit**

Worldwide, there has been a renewed interest in natural products. This interest is a result of factors such as: consumer's belief that natural products are superior; consumer's dissatisfaction with conventional medicines; changes in laws allowing structure-function claims which results in

more liberal advertising; aging baby boomers; national concerns for health care cost. The potential for developing antimicrobials into medicines appears rewarding, from both the perspective of drug development and the perspective of phytomedicines. The immediate source of financial benefit from plants based antimicrobials is from the herbal products market.

## Literature Review

### **Anthelmintic activity:**

The roots of *K. pinnata* were subjected to petroleum ether, chloroform, methanol and aqueous solvent respectively for extraction and the *in-vitro* evaluation of anthelmintic activity was done against *Pheretimaposthuma* (Annelida) and *Ascardiagalli* (nematode). The results reveal that chloroform, methanolic and aqueous extract of *K. pinnata* have significant anthelmintic activity while petroleum ether does not show any activity against helminth. Methanolic extract of root of *K. pinnata* was found to be most effective as an anthelmintic as compared to other. The roots extract of *K. pinnata* not only demonstrated paralysis but also caused deaths of worms especially at higher concentrations of 100 mg/ml, in shorter time as compared to the reference drug, Piperazine citrate. Phytochemical analysis of the crude extracts revealed the presence of tannins which were shown to produce anthelmintic activity (Majaz et al., 2011).

### **Immunosuppressive effect:**

The aqueous extract of *K. pinnata* leaves was found to cause significant inhibition of cell-mediated and humoral immune responses in mice. The spleen cells of animals pre-treated with *K. pinnata* showed a decreased ability to proliferate in response to both mitogen and to antigen *in vitro*. Treatment with *K. pinnata* also impaired the ability of mice to mount a delayed-type hypersensitivity reaction (DTH) to ovalbumin. The intravenous and topical routes of administration were the most effective by almost completely abolishing the DTH reaction. The intraperitoneal and oral routes reduced the reaction by 73 and 47% of controls, respectively. The specific antibody responses to ovalbumin were also significantly reduced by treatment. Together, these observations indicate that the aqueous extract of *K. pinnata* possesses an immunosuppressive activity (Bergmann et al., 2006).

**Wound healing activity:**

The extract of *K. pinnata* was evaluated for its wound healing activity by using excision wound model in rats. On the 11th day wounding, there was a significant increase in the wound-healing activity in the animal treated with *K. pinnata* ethanolic extract compared to animals which received the control treatment and standard treatment. Significant progressive reduction in the wound area was observed by day 11 (86.3%) when compared to the control (68.0%) and standard (85.5%). The histological analysis showed that *K. pinnata* leaf extract exhibited significant wound healing potential. The wound healing exhibited by the extract may be attributed to the presence of steroid glycosides. The medicinal plant has been shown to have a significant quantity of bufadienolide, a steroidal aglycone which exists in the plant as steroidal glycoside (Nayak et al., 2010).

**Antihypertensive activity:**

The effects of aqueous leaf extract of *K. pinnata* on the blood pressure of anaesthetized cats as well as on the liver and kidney status of the rabbit were investigated in this study. The results revealed that the extract produced a small fall in the blood pressure of the anaesthetized cat and also reduced the effect of adrenaline-induced elevation of blood pressure. It was concluded that the pharmacological basis for the use of *K. pinnata* among the Igbos of Nigeria to lower blood pressure was established by this study. However, the facts that the reduction in blood pressure produced is slight and the *K. pinnata* leaf extract is potentially organotoxic which negates its use as a blood pressure lowering agent (Ghasi et al., 2011).

**Hepatoprotective activity:**

Juice of the fresh leaves is used very effectively for the treatment of jaundice in folk medicines of Bundelkhand region of India. The juice of the leaves and the ethanolic extract of the marc left after expressing the juice were studied in rats against CCl<sub>4</sub>-induced hepatotoxicity. The test material was found effective as hepatoprotective as evidenced by *in vitro*, *in vivo* and histopathological studies. The juice was found to be more effective than ethanolic extract (Yadav and Dixit, 2003).

**Antinociceptive, anti-inflammatory and antidiabetic activity:**

In order to scientifically appraise some of the ethnomedical uses of *K. pinnata* leaves, a study was undertaken to investigate the antinociceptive, anti-inflammatory and antidiabetic properties of the plant's leaf aqueous extract in experimental animal models. *K. pinnata* leaf aqueous extract (BPE, 25 to 800 mg/kg i.p.) produced significant ( $P < 0.05$  to 0.001) antinociceptive effects against thermally- and chemically-induced nociceptive pain stimuli in mice. The plant extract (BPE, 25 to 800 mg/kg p.o. or i.p.) also significantly ( $P < 0.05$  to 0.001) inhibited fresh egg albumin-induced acute inflammation and caused significant ( $P < 0.05$  to 0.001) hypoglycaemia in rats. The results of this experimental animal study suggest that *K. pinnata* leaf aqueous extract possesses antinociceptive, anti-inflammatory and hypoglycaemic properties. The different flavonoids, polyphenols, triterpenoids and other chemical constituents of the herb are speculated to account for the observed antinociceptive, anti-inflammatory and antidiabetic properties of the plant (Ojewole, 2005).

**Nephroprotective and antioxidant activity:**

Harlalka et al. (2007) evaluated the aqueous extract of *K. pinnata* for its protective effects on Gentamycin-induced nephrotoxicity in rats. It was observed that the aqueous extract of *K. pinnata* leaves significantly protects rat kidneys from Gentamycin-induced histopathological changes. Gentamycin-induced glomerular congestion, peritubular and blood vessels congestion, epithelial desquamation, accumulation of inflammatory cells and necrosis of the kidney cells were found to be reduced in the group receiving the leaf extract of *K. pinnata* along with Gentamycin. Urine creatinine, serum creatinine, blood urea, blood urea nitrogen and the weights of the kidneys were found to be significantly increased in rats treated with only Gentamycin; whereas the treatment with the aqueous extract of *K. pinnata* was found to protect the rats from such effects of Gentamycin. The volume of urine was found to be significantly increased in the rats treated with *K. pinnata* leaf extract. In case of histopathological examination, control rats showed normal glomerular and tubular histology whereas Gentamycin was found to cause glomerular, peritubular and blood vessel congestion and result in the presence of inflammatory cells in kidney sections from the Gentamycin-treated group. Concurrent treatment with the extract was found to reduce such changes in kidney histology induced by Gentamycin. *In-vitro* studies revealed that the *K. pinnata* leaf extract possesses



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significant antioxidant as well as oxidative radical scavenging activities. Quercetin and kaemferol have been detected in the leaves of *K. pinnata* (Harlalka et al., 2007). Morales et al. (2006) suggested that quercetin has a marked protective effect on cadmium-induced nephrotoxicity that results from an increase in Metallothionein, a small cysteine-rich protein and eNOS (endothelial nitric oxide synthase) expression and the inhibition of COX-2 (cyclooxygenase-2) and iNOS (inducible nitric oxide synthase) expression.

### **Antimicrobial activity:**

The roots of *K. pinnata* were subjected to petroleum ether, chloroform, methanol and aqueous solvent respectively for extraction and *in vitro* evaluation of antimicrobial activity was done against *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa* and *Candida albicans*. Methanolic extract of roots of *K. pinnata* was found to be most effective as antibacterial as compared to others while none of the extracts showed activity against *C. albicans* (Quazi et al., 2011). Akinpelu (2000) in a study found that 60% methanolic leaf extract inhibits the growth of five out of eight bacteria used at a concentration of 25 mg/ml. *Bacillus subtilis*, *E. coli*, *Proteus vulgaris*, *Shigella dysenteriae*, *S. aureus* were found to be inhibited while *Klebsiella pneumoniae*, *P. aeruginosa* and *C. albicans* were found to resist the action of the extract. Chemical investigation of the bioactive constituents from the leaf of *K. pinnata* resulted in the isolation of two new novel flavonoids; 5I Methyl 4I,5, 7 trihydroxyl flavone and 4I, 3, 5, 7 tetrahydroxy 5-methyl 5I-propenamine anthocyanidines. The antimicrobial observation of the aforementioned compounds could be responsible for the activity of *K. pinnata* and its use in herbal medicine in Nigeria (Okwu and Nnamdi, 2011).

### **Analgesic and anticonvulsant effects:**

The analgesic effect of methylene chloride/methanol (1:1) (CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH) extract and its hexane, methylenechloride (CH<sub>2</sub>Cl<sub>2</sub>), ethyl acetate, *n*-butanol fractions and aqueous residue was evaluated using acetic acid, formalin and pressure test. The anticonvulsant effects of the CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH extract were also investigated on seizures induced by pentylenetetrazol (PTZ), strychnine sulphate (STN) and thiosemicarbazide (TSC). CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH extract and its fractions administered orally exhibited protective effect of at least 30% on the pain induced by acetic acid. The CH<sub>2</sub>Cl<sub>2</sub> fraction at 300 mg/kg showed a maximal effect of 78.49%.

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The CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH extract and its CH<sub>2</sub>Cl<sub>2</sub> fraction at the doses of 150 and 300 mg/kg significantly reduced the first phase of pain induced by formalin while the second phase was completely inhibited. The CH<sub>2</sub>Cl<sub>2</sub> fraction produced more than 45% reduction in the sensitivity to pain induced by pressure. The CH<sub>2</sub>Cl<sub>2</sub>/CH<sub>3</sub>OH extract of *K. pinnata* significantly increased the latency period in seizures induced by PTZ and significantly reduced the duration of seizures induced by the three convulsant agents. The extract protected 20% of animals against death in seizures induced by TSC and STN. These results suggest a peripheral and central analgesic activities as well as an anticonvulsant effect of the leaves of *K. pinnata* (Nguelefack et al., 2006)

### **Leishmaniasis activity:**

Muzitano et al. (2009) carried out an investigation to study the effect of *K. pinnata* on cutaneous leishmaniasis. In order to demonstrate the safety and oral activity of *K. pinnata*, different flavonoids were extracted from the plants and were evaluated *in vivo* in murine model of cutaneous leishmaniasis. Daily oral doses of quercetin 3-O- $\alpha$ -L-arabinopyranosyl,  $\alpha$ -L-rhamnopyranoside, quercetin 3-O- $\alpha$ -L-rhamnopyranoside and free quercetin (16 mg/kg body weight) were administered. It was observed that they were able to control the lesion growth caused by *Leishmania amazonensis* and significantly reduce the parasite load. These flavonoids were as effective as the crude *K. pinnata* aqueous extract given at 320 mg/kg body weight. HPLC-DAD-MS analysis of the plasma of extract-treated mice suggested that quercetin and quercetin glucuronides are the main metabolites of *K. pinnata* quercetin glycosides. These results indicate that quercetin glycosides are important active components of the aqueous extract and that they possess potent oral efficacy against cutaneous leishmaniasis.

### **Diuretic and anti-urolithiatic activity:**

Patil et al. (2009) studied the diuretic and anti-urolithiatic activity of *K. pinnata*. Hydroalcoholic extract of leaves of *K. pinnata* was administered to male Wistar rats orally and intraperitoneally. The effect of the extract on urine output was determined by comparing the urine volume collected by keeping the individual animals in metabolic cages. Calcium oxalate urolithiasis was induced in rats by giving ethylene glycol orally for 7 days and the effect of the extract was observed by its concurrent administration. The extract was found to have significant diuretic and

### Neuropharmacological Evaluation of *k. pinnata*

anti-urolithiatic activity and the intraperitoneal administration of the extract gave more potent diuretic effect.

#### **Anti-tumor activity:**

Five bufadienolides (1-5) isolated from the leaves of *K. pinnata* were examined for their inhibitory effects on Epstein-Barr virus early antigen (EBV-EA) activation in Raji cells induced by the tumor promoter, 12-O-tetradecanoylphorbol-13-acetate. All bufadienolides showed inhibitory activity, and bryophyllin A (1) exhibited the most marked inhibition ( $IC_{50} = 0.4$  microM) among the tested compounds. Bryophyllin C (2), a reduction analogue of 1, and bersaldegenin-3-acetate (3) lacking the orthoacetate moiety were less active. These results strongly suggest that bufadienolides are potential cancer chemopreventive agents (Supratman et al., 2001).

#### **Anti-allergic activity:**

Cruz et al. (2008) reported on the protective effect of *K. pinnata* in fatal anaphylactic shock, likewise a Th2-driven immunopathology and the identification of its active component. *In vitro*, *K. pinnata* prevented antigen-induced mast cell degranulation and histamine release. Oral treatment with the quercitrin flavonoid isolated from the plant prevented fatal anaphylaxis in 75% of the animals. These findings indicate that oral treatment with *K. pinnata* effectively down-modulates pro-anaphylactic inducing immune responses. Protection achieved with quercitrin, although not maximal, suggests that this flavonoid is a critical component of *K. pinnata* extract against this extreme allergic reaction.

#### **Neuropharmacological activity:**

Effects of aqueous leaf extracts of *K. pinnata* on some neuropharmacological activities were studied in mice. The extract was found to produce a profound decrease in exploratory activity in a dose-dependent manner. It also showed a marked sedative effect as evidenced by a significant reduction in gross behaviour and potentiation of pentobarbitone-induced sleeping time. It delayed onset in strychnine- and picrotoxin-induced convulsion (seizures) respectively with the protective effect being significantly higher in picrotoxin- than strychnine-induced convulsion. It

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also decreases the rate of picrotoxin-induced mortality in mice with LD<sub>50</sub> of 641 mg/kg. The totality of these effects showed that the extract possesses depressant action on the central nervous system (Salahdeen and Yemitan, 2006).

#### **Antimutagenic activity:**

Plant has potent antihistamine and anti-allergic activity. The methanolextract of the leaves has also been reported to have histamine receptor (H<sub>1</sub>) antagonism in the ileum, peripheral vasculature and bronchial muscle and protect against chemically induced anaphylactic reactions and death by selectively blocking histamine receptors in the lungs. Quercetin-3-O- $\alpha$ -L-rabinopyranosyl (1 $\rightarrow$ 2)- $\alpha$ -L-rhamnopyranoside showed anti-allergic activity in rats. Obaseiki-Ebor *et al* investigated that organic solvent extracts of leaves had inhibitory activity for His<sup>-</sup> to His<sup>+</sup> reverse-mutations induced by ethyl methanesulfonate acting on *S. typhimurium* TA100 or TA1002 and were also active against reversions induced by 4-nitro-o-phenylenediamine and 2-aminofluorene in TA98. The alkaloidal/ water soluble and acid fraction had no appreciable antimutagenic activity.

#### **Anti-ulcer activity:**

Adesanwoet *al* in his study showed a significant reduction in incidence of ulceration and mean basal and histamine-stimulated gastric acid secretion in a dose-dependent manner thus justifying its use as an anti-ulcer agent in folklore medicine.

#### **Uterine Contractility:**

B. Gwehenberger *et al* characterise the phytotherapeutic tocolytic effect of *B. Pinnatum* in vitro versus the conventional betamimetic, fenoterol, in human myometrium.

Contractility was measured in strips of term myometrium biopsied at caesarean section in 14 women and exposed to increasing concentrations of *B. Pinnatum* versus +/- oxytocin 1 U/l. Result state inhibition of spontaneous contraction was concentration dependent. *B. Pinnatum* increased contraction frequency by 91% at constant amplitude and inhibited oxytocin-stimulated contractions by 20% at constant amplitude with slightly decreased frequency. Fenoterol decreased contraction by 50% with a significant decrease in frequency.

**Toxic to cattle:**

Mckenzie *et al* investigated that cardiac glycoside poisoning was produced in calves given flower heads of the hybrid *Bryophyllum Species* and found that for each plant (except *B. tubiflorum*), 2 calves were each given a single dose of 20 g wet weight per kg bodyweight. The results of the calf toxicity experiment with the amounts of bufadienolide measured in the plants suggests that bryotoxins A, B and C probably account for the observed disease.

**Anticancer:**

Bryophyllin compounds have marked anticancer therapeutic value against cancer cells<sup>15</sup>. Bersaldegenin-1, 3, 5-orthoacetate inhibited cancer cell growth on several cancer lines.

**Insecticidal, Fungitoxic and Phytotoxic activity:**

Alabiet *et al* studied to evaluate the fungitoxic and phytotoxic effects of extracts on the fungal pathogens inducing wilting on cowpea grown in Ago-Iwoye, South Western Nigeria. The extract reduces the Disease Infection Rate (DIR) in treated plants. *Sclerotium rolfsii* induced wilting of between 4 and 12% on cowpea seedlings treated with plant extract under field conditions while about 39.6% incidence of cowpea seedlings wilting was observed under control experiment on the same experimental plot. The extracts increased significantly the plant height, shelf life, relative water content and chlorophyll contents of the cowpea seedlings during both the wet and dry season. On the other hand, the extracts significantly reduced transpiration rate and stomata aperture of treated plant in both seasons. Furthermore, application of these extracts on the cowpea plants significantly enhanced the Leaf Area Index (LAI), number of branches and pods per plant, total dry matter per plant, weight per pod, 100 grains weight and grain yield in both season. The extracts also inhibited the release of current photosynthates from treated plants thus maintaining the water status of plant and also making photosynthates which can be oxidized to release energy needed for growth available to treated plants.

**Clinical usage**

- The leaves are useful in burns, boils, bites of insects, congestive ophthalmia dysuria, diarrhoea, dysentery, impetigo, polyuria, plegmon, swellings, tuberculosis, ulcers and wounds

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- The leaf juice 3 g, jeera 3 g and ghee 6 g is mixed and given for blood mixed diarrhoea
- The leaf poultice is applied on wounds, sprains, swellings and inflammations
- The leaf juice is useful in cholera
- The leaf juice mixed with Kali Mirch is useful in blood oozing piles and haemorrhoids
- The leaf powder with Kali Mirch is also useful in inflammation, burning in urination and blockedurination and leprosy
- The leaves roasted over fire are applied to places of wounds and surgical sutures in the skin to prevent discoloration of the skin

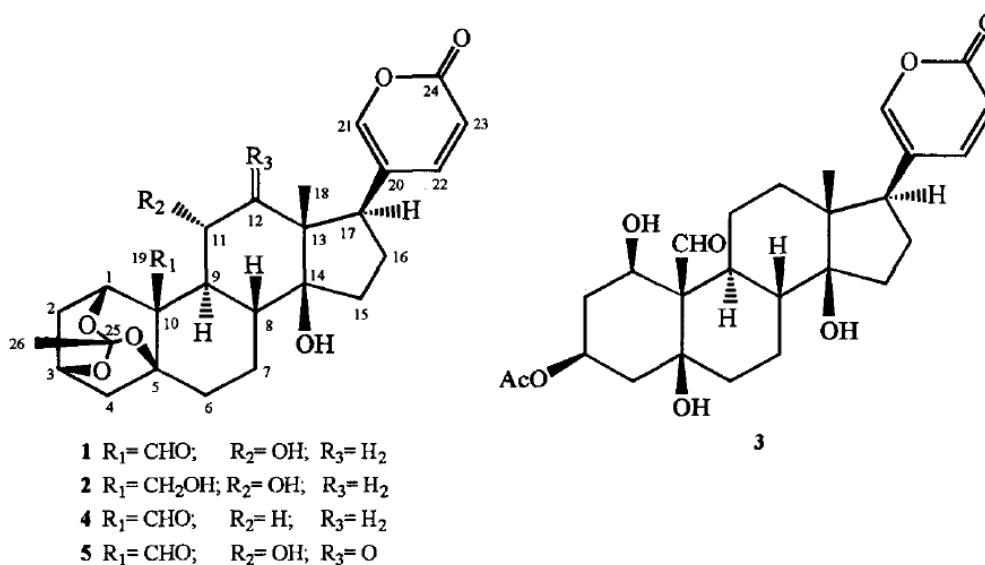
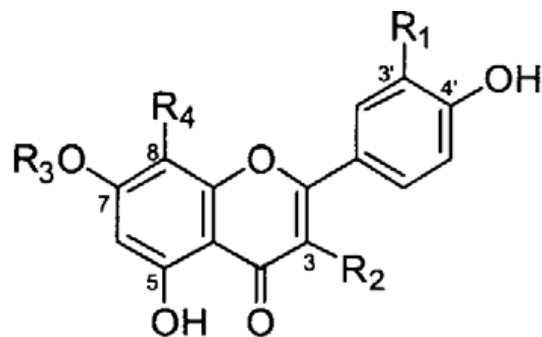
**Chemical Constituents Isolated From KalanchoePinnata:**

Fig. Structures of Compounds 1-5.



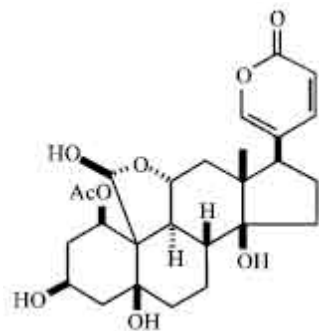


Fig 1. Bryophyllin

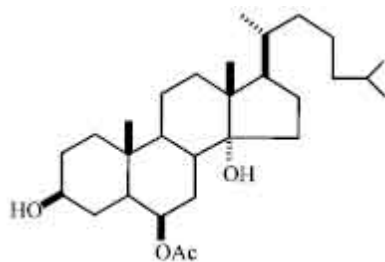


Fig 2 Bryophyllol

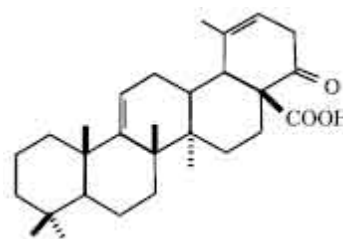


Fig 3 Bryophollone

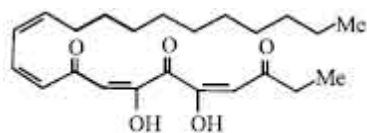
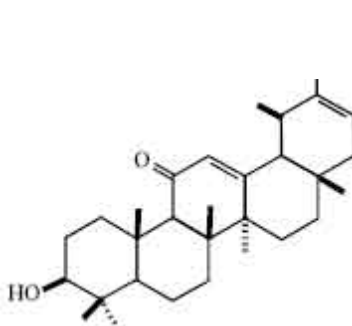


Fig 5 Bryophynol

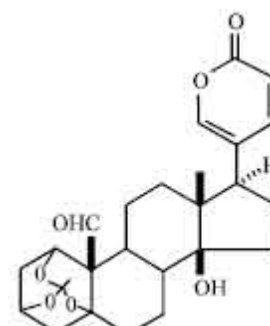


Fig 6 Bersaldegenin

Fig 4 Bryophollenone

**Chemical constituents:** *B. Pinnatum* is rich in alkaloids, triterpenes, glycosides, flavonoids, cardenolides, steroids, bufadienolides and lipids. The leaves contain a group of chemicals called bufadienolides which are very active. Bufadienolides like bryotoxin A, B, C which are very similar in structure and activity as two other cardiac glycosides, digoxin and digitoxin and possess antibacterial, antitumor, cancer preventative and insecticidal actions. Bufadienolides- Bryophyllin A (bryotoxin); Bryophyllin B (Fig. 1); Bryophyllol (Fig. 2); Bryophollone (Fig. 3); Bryophollenone (Fig. 4); Bryophynol (Fig. 5).

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**Phenols, Phenylpropanoids and Flavanoids:** Syringic acid, caffeic acid 10 , 4-hydroxy-3-methoxy-cinnamic acid, 4-hydroxybenzoic acid, p-hydroxycinnamic acid, paracoumaric acid, ferulic acid, protocatechuic acid, phosphoenolpyruvate, protocatechuic acid isolated from aerial parts of plants. Leaves contains astragalin, 3,8-dimethoxy-4, 5, 7-trihydroxyflavone, friedelin, epigallocatechin-3-osyringate, luteolin, rutin, kaempferol , quercetin , quercetin-3L-rhamnoside-L-arabino furanoside ;quercetin-3-Odiarabinoside, kaempferol-3-glucoside , kaempferol-3-O- $\alpha$ -L-arabinopyranosyl (1 $\rightarrow$ 2)  $\alpha$  - L-rhamnopyranoside, quercetin-3-O- $\alpha$ -L-arabinopyranosyl (1 $\rightarrow$ 2) $\alpha$  -L-rhamnopyranoside and 4',5-dihydroxy-3',8-dimethoxy flavone-7O- $\beta$ -D-glucopyranoside. Because of its restricted occurrence and its abundance in *B. Pinnatum*, flavonoid may be a chemical marker of the plant of high therapeutic potential.

**Triterpenoids and Steroids:** The plant contains  $\alpha$ -amyirin,  $\alpha$ -amyirinacetate,  $\beta$ -amyirin,  $\beta$ -amyirinacetate, bryophollenone , bryophollone , taraxerol,  $\Psi$ -taraxasterol , pseudo taraxasterol, 18- $\alpha$ -oleanane, friedelin, glutinol. The cardienolide and steroidal contents includes  $\beta$ -sitosterol , bryophyllol, bryophynol , bryophyllinB (Antitumor) , bryophyllin A (bryotoxin C , bufadienolide1, 3, 5-orthoacetate) with potent cytotoxicity, a insecticidal bufadienolide bryophyllin C and bersaldegenin-3-acetate , bryotoxinA, bryotoxin B , bersaldegenin-1, 3, 5-orthoacetate, campesterol , 24-ethyl-25-hydroxycholesterol, isofucosterol , clionasterol, codisterol , peposterol, 22-dihydrobrassicasterol, clerosterol, 24-epiclerosterol, 24ethyl- desmosterol, 25-methyl-5 $\alpha$ -ergost-24 -en-3- $\beta$ -ol, ergosta-5-24 -dien-3- $\beta$ -ol, 25-methyl-ergosta-5-24 -dien-3- $\beta$ -ol, 5 $\alpha$ -stigmast-24-en-3- $\beta$ -ol , (24s)-stigmast-25-en-3- $\beta$ -ol, (24r)- 5 $\alpha$ -stigmasta-7-25-dien-3- $\beta$ -ol, (24s)-5 $\alpha$ -stigmasta-7,25 dien-3- $\beta$ -ol, 24(R)-stigmasta-5,25-dien-3- $\beta$ -ol, stigmasterol , patuletin , 3-O-(4-O-acetyl- $\alpha$ -L-rhamnopyranosyl)-7O-(2-O-acetyl- $\alpha$ -L-rhamnopyranoside) patuletin, 3-O- $\alpha$ -L-rhamno pyranosyl-7-O-(2-O-acetyl- $\alpha$ -L-rhamnopyranoside) patuletin, 3-O-(4-O-acetyl- $\alpha$ -L-rhamnopyranosyl)-7-O-rhamnopyranoside patuletin are isolated from aerial parts 17,18 .

**Fatty Acids, Minerals and Others:** Fatty acid fraction includes palmitic acid (89.3%), stearic acid (10.7%), traces of arachidic and behenic acid . Plant also contains HCN, oxalic acid , citric acid, isocitric acid, oxaloacetate, malic acid and succinic acid. The plant is rich in vitamins and amino acids; ascorbic acid, riboflavin, thiamine, niacin, pyridoxine, glycine, cysteine, casein



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hydrolysate, glutamic acid, protein hydrolysate, methionine, tyrosine, phenylalanine 19. Food contents are carbohydrates, protein, lipids, acids, iodine. The herb is good source of mineral elements such as Na, Ca, K, P, Mg, Mn, Fe, Cu, Zn. Sugar contents includes raffinose, lactose, sucrose, glucose, galactose, fructose. Plant also contains alkaloids, tannins, phenanthrene derivatives: 2(9-decenyl)-phenanthrene, 2(9-undecenyl)-phenanthrene, alkanes (C 25-35 ), alkanols (C 26-34 ), ntriacontane, hentriacontane<sup>20</sup>.

## Materials and Method

### Plant material and extraction:

The leaves of *Kalanchoepinnata* were obtained from different places in Dhaka, Bangladesh in August, 2009. The plant was taxonomically identified by Mr. Manzur-ul-Kadir Mia, ex-Principal Scientific Officer and Curator of Bangladesh National Herbarium at Dhaka. The dried leaves of *Kalanchoepinnata* (leaves were air-dried in the shade for 144 hours) were grounded into a fine powder and were extracted with methanol at a ratio of 1:5 (w/v). After 24 hours, the mixture was filtered; filtrate was collected and the residue was again extracted with methanol at a ratio of 1:3 (w/v) for 24 hrs. Filtrates were combined and evaporated to dryness using rotary evaporator.

### Chemicals

Glacial acetic acid was obtained from Sigma Chemicals, USA; diazepam was obtained from Square Pharmaceuticals Ltd., Bangladesh. All other chemicals were of analytical grade.

### Experimental design:

The animals were divided into five groups. The first groups of animals were used for the behavioural changes and acute toxicity studies. The second groups were used for the exploratory behaviour. The third groups were used for the benzodiazepam sleeping time. The fourth group were used also for the muscle relaxant tests. (Grp IV). The group which is group five was used for the study of anticonvulsant activity of the aqueous extract. Each group of animals was sub-divided into control and experimental groups, which was treated with the extract at sub-lethal doses of 50, 100, 200mg/kg bodyweight while the control received normal saline. The vehicle and the extracts were administered intraperitoneally before the experimentation.

### Materials & Reagents:

1. Rotary evaporator ( IKA RV05 basic, Biometra)
2. Closed test tube
3. beaker

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4. Aluminium foil paper
5. spatula
6. pipette
7. Blender
8. Sodium Chloride
9. Distilled Water
10. Beaker
11. Cotton buds
12. Glass container for Isotonic Solution
13. Gas burner
14. Forceps
15. Labels
16. filter paper (Whatman 40)
17. permanent marking pen
18. aluminium foil paper
19. Plant extracts
20. 50 ml beakers
21. Electric balance (ELH 3000, Shimadzu, Japan)
22. pipette pumper
23. scissor
24. Ethyl acetate
25. n-Hexane
26. Acetic acid
27. Methanol
28. Dichloromethane
29. Sulfuric acid
30. Capillary tube
31. Pencil
32. Scale

## Test Methods

### Acetic Acid Induced Writhing Test:

The animals randomly divided into four groups consisted of 5 mice in each group. Group- I was kept as control giving 1% Tween-80 in distilled water (10ml/kg b.w). Group II received Diclofenac-Na as standard drug at a dose of 10mg/kg of body weight (p.o). Group-III and Group-IV received the test compound acetic acid extract of *Kalanchoepinnata* at the doses of 200 mg/kg and 400 mg/kg of body weight (p.o) respectively. Thirty minutes after treatment, the mice were given an intraperitoneal (i.p.) injection of 0.7% acetic acid in a volume of 10ml/kg to induce the specific contraction of the body referred to as "writhing". The no. of writhings occurring between 5 and 15 minutes after acetic acid injection was recorded. *Kalanchoepinnata* leaves was examined using method as previously described (Deb *et al.*, 2010) with minor modifications. In the writhing test, pain was induced through intraperitoneal administration of 1% acetic acid at a dose of 10 ml 1% acetic acid/kg body weight. For experiments with *Kalanchoepinnata* leaves, mice were separated into five groups of five mice each. Group-I served as control and was administered vehicle (1% Tween 80 in water, 10 mg/kg body weight).

### Formalin induced licking response model

#### Study design

Experimental animals were randomly selected and divided into three groups denoted as group-I, group-II and group-III consisting of 5 mice in each group. Each group received a particular treatment i.e. control, positive control and a dose of the *k. pinnata* extract (Test group). Each animal was weighed properly and the dose of the test samples and control materials were adjusted accordingly.

#### Methodology

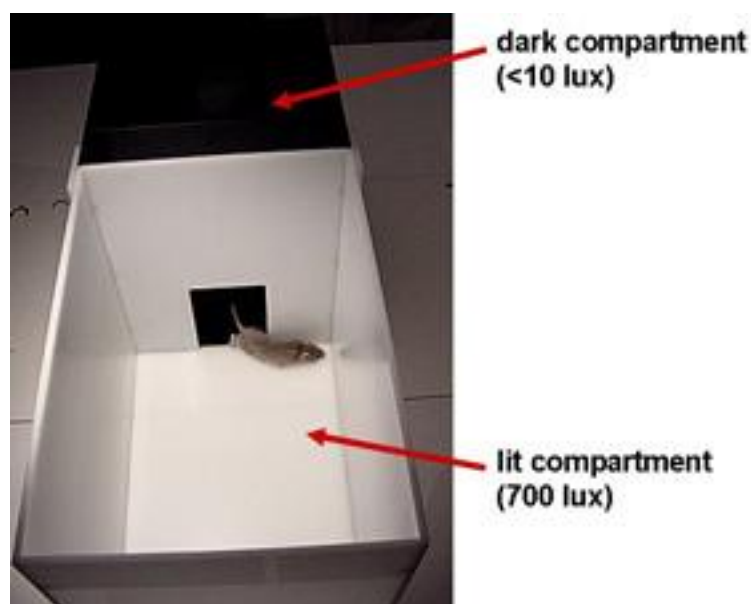
Acetic acid induced pain model was performed according to the method described by Sharma *et al.*, (2010). 20µl of 1.0% v/v acetic acid was injected subcutaneously into the right hind paw of

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mice. The time (in sec) spent in licking the paw and the biting responses of the injected paw were taken as an indicator of pain response. The rats were observed for 30 min after the injection of acetic acid, and the amount of time spent licking the injected hind paw was recorded. The first 5 min post acetic acid injection is referred to as the early phase and the period between 15 and 30 min as the late phase. Extract (200 mg/kg, orally) and diclofenac sodium (0.5 mg/kg, i.p) were administered 30 min prior to formalin injection. Control animals received 10 ml/kg of distilled water, orally.

### Hole cross test:

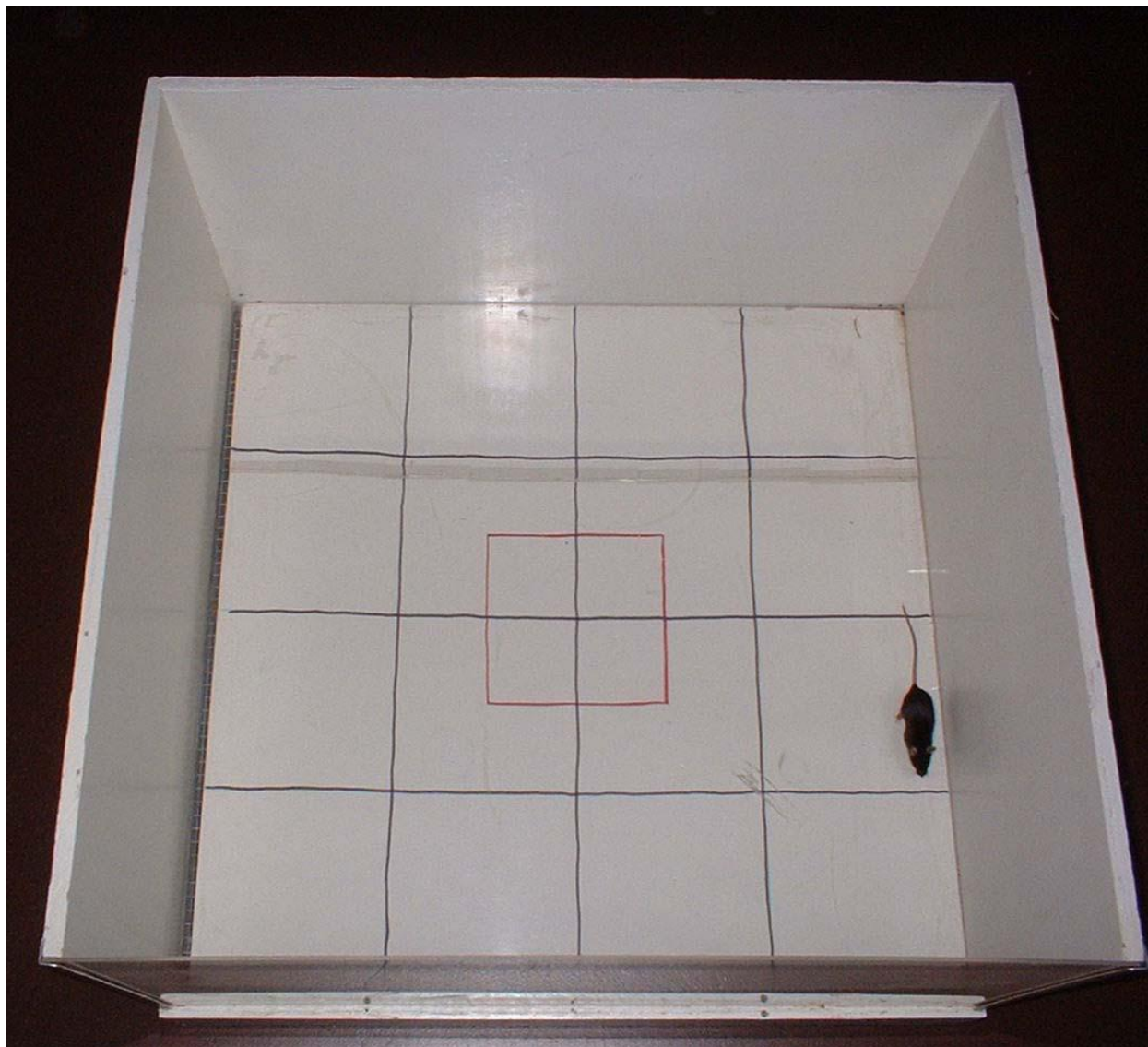
The method was carried out as described by Takagi *et al.* (8). A steel partition was fixed in the middle of a cage of 30 × 20 × 14 cm. A hole of 3 cm diameter was made at a height of 7.5 cm in the center of the cage. The animals were divided into control, positive control, and test groups containing five mice each. The test groups received acetic acid extract of *kolanchoepinnata* at doses of 200 and 400 mg/kg body weight orally whereas the control group received vehicle (1% Tween 80 in water). The number of passages of a mouse through the hole from one chamber to the other was counted for a period of 3 min at 0, 30, 60, 90, and 120 min after oral administration of both doses of the test drug.



**Figure: Hole test**

### **Open field test:**

In the open field test, the animals were divided into control, positive control, and test groups containing five mice each. The test groups received acetic acid extract of the leaves of *chalanchoe pinnata* at doses of 400 mg/kg body weight orally whereas the control group received vehicle (1% Tween 80 in water). The floor of a half square meter open field (9,10) was divided into a series of squares each alternatively colored black and white. The apparatus had a 40 cm height wall. The number of squares visited by the animals was counted for 3 min at 0,30, 60, 90, and 120 min after oral administration of both doses of the test drug.

**Figure: Open field test**

The open field can be of different sizes; small (38 x 38 cm), or large (72 x 72 cm). The small open field can also serve as a hole board and as a test chamber for the novel object recognition task. The large open field is used for measuring anxiety and exploration as well as locomotion as it has a large center arena.

**Apparatus**

The open field apparatus was constructed of white plywood and measured apparatus. Blue lines were drawn on the floor with a marker and were visible through the clear Plexiglas floor. The lines divided the floor into sixteen 18 x 18 cm squares. A central square (18 cm x 18 cm) was drawn in the middle of the open field (Brown, Corey, & Moore, 1999). The central square is used because some mouse strains have high locomotor activity and cross the lines of the test

## Neuropharmacological Evaluation of *k. pinnata*

chamber many times during a test session. Also, the central square has sufficient space surrounding it to give meaning to the central location as being distinct from the outer locations (Carrey, McFadyen, & Brown, 2000).

The maze was located in a 1.8 x 4.6 m test room and lit by a 60-watt red lamp for background lighting. The open field maze was cleaned between each mouse using 70 % ethyl alcohol. Behavior was scored with Hindsight for MS-dos (ver 1.5), and each trial was recorded for latter analysis, using a video camcorder (Hitachi, VM-7500LA) positioned above the apparatus. Measures of line crosses were obtained with an automated camera-based computer tracking system (Limelight, Actimetrics) on an IBM PC computer with the camera fixed to the ceiling, 2.1 m above the apparatus.

### **Procedure**

Mice were carried to the test room in their home cages and were handled by the base of their tails at all times. Mice were placed into the center, or one of the four corners of the open field and allowed to explore the apparatus for 5 minutes. After the 5 minute test, mice were returned in their home cages and the open field was cleaned with 70 % ethyl alcohol and permitted to dry between tests. To assess the process of habituation to the novelty of the arena, mice were exposed to the apparatus for 5 minutes on 2 consecutive days.

### **Behaviours scored**

The behaviours scored (Brown et al, 1999) included:

1. Line Crossing: Frequency with which the mice crossed one of the grid lines with all four paws.
2. Center Square Entries: Frequency with which the mice crossed one of the red lines with all four paws into the central square.
3. Center Square Duration: Duration of time the mice spent in the central square.
4. Rearing: Frequency with which the mice stood on their hind legs in the maze.
5. Stretch Attend Postures: Frequency with which the animal demonstrated forward elongation of the head and shoulders followed by retraction to the original position.
6. Grooming: Duration of time the animal spent licking or scratching itself while stationary.
7. Freezing: Duration with which the mouse was completely stationary.
8. Urination: number of puddles or streaks of urine.



9. Defecation: number of fecal boli produced.

Each animal was then given a score for total locomotor activity that was calculated as the sum of line crosses and number of reaction.

**Why open field test is important:**

The Open Field Test (Walsh & Cummins, 1976) provides simultaneous measures of locomotion, exploration and anxiety. The number of line crosses and the frequency of rearing are usually used as measures of locomotor activity, but are also measures of 4

## Neuropharmacological Evaluation of *k. pinnata*

exploration and anxiety. A high frequency of these behaviours indicates increased locomotion and exploration and/or a lower level of anxiety. The number of central square entries and the duration of time spent in the central square are measures of exploratory behaviour and anxiety. A high frequency/duration of these behaviours indicates high exploratory behaviour and low anxiety levels.

Stretch attend postures are “risk-assessment” behaviours which indicate that the animal is hesitant to move from its present location to a new position (Blanchard, Griebel & Blanchard, 2001) and thus a high frequency of these postures indicates a higher level of anxiety. Grooming behaviour is a displacement response and is expected to be displayed in a novel environment (Espejo, 1997). Therefore grooming behaviour should decrease with repeated exposure to the testing apparatus. Defecation and urination are often used as measures of anxiety, but the validity of defecation as a measure of anxiety has been questioned (Lister, 1990). Hall (1934) describes defecation and urination as indices of anxiety in rodents. He argues that the animal will have reduced locomotion in a novel environment but the autonomic nervous system will be activated which will increase defecation in this noxious arena. However, Bindra & Thompson (1953) argue that there is no significant relation between fearfulness and urination and defecation as measured in the Open Field test. Nevertheless, Bindra & Thompson (1953) agree that defecation and urination in a novel environment are signs of emotionality, which is not to be equated with fearfulness or timidity.

### **Factor Analysis**

Several dependent variables measured in the open field correlate significantly with one another, such as: line crosses and rearing, as well as line crosses and central square activity (Walsh & Cummins, 1976). Factor analysis of open field behaviour generally yields 3 factors but the names of these factors vary. Jahkel et al. (2000) identified 3 factors: **Activity** (53.5% of the variance) (line crosses, time active); **Exploration** (15.5% of the variance) (Center squares crossed, time in center); and **Irritation** (9.5% of the variance) (time passive). Ramos et al. (1997) also identified 3 factors: **Anxiety or Approach/Avoidance** (36.6% of variance) (locomotion in center squares); **Locomotor Activity** (30.3% of variance) (total lines crossed, lines crosses in outer squares) and **Defecation** scores (18.3% of variance). Crusio & Schwegler (1987) also found 3 factors: **Locomotion** (rearing, lines crossed); **Grooming** (grooming frequency and duration); and **Defecation** (sniffing, defecates). These studies did not always

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measure the same behaviours and the different factors identified indicate that the definitive study of the open field test has yet to be conducted.

### **Repeated Exposure**

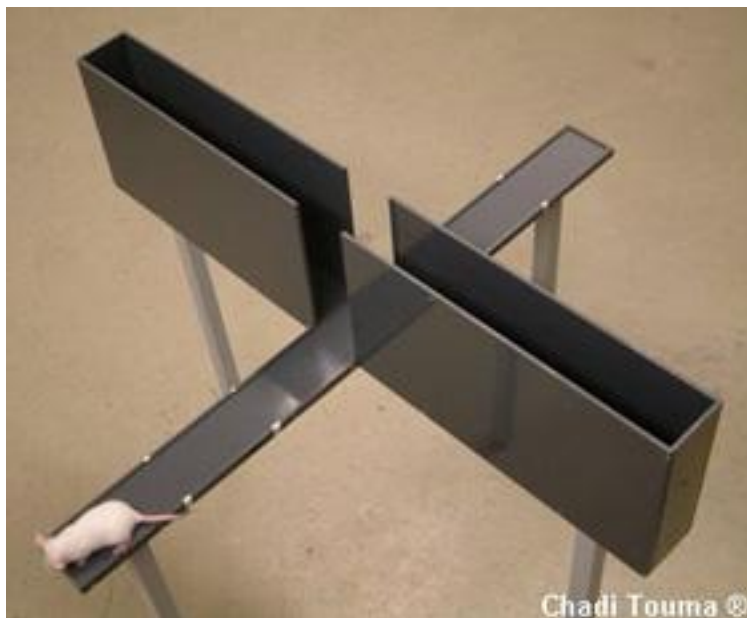
Repeated exposure to the open field apparatus results in time dependent changes in behaviour (Choleris, et al., 2001). At first, when the apparatus is novel to the animals, more fear-related behaviours (such as stretch attends and activity in the corners and walls of the open field) are displayed. However, with repeated trials, more exploration and locomotor activity (such as rearing and line crosses as well as more central square activity) is observed. There are, however, strain differences in behaviour after repeated testing in the open field. With repeated exposure, some strains show increased activity while others show habituation and decreased activity levels and others show no change (Bolivar et al. 2000).

### **EPM test**

The method initially suggested by Handley and Mithaniwas employed with minor modifications (12). The apparatus consists of two open arms (5 × 10 cm) and two closed arms (5 × 10 × 15 cm) radiating from a platform (5 × 5 cm) to form a plus-sign figure. The apparatus was situated 40 cm above the floor. The open arms edges were 0.5 cm in height to keep the mice from falling and the closed-arms edges were 15 cm in height. Sixty minutes after administration of the test drugs, each animal was individually placed in the center of the EPM and were allowed 5 min for free exploration. Next, the number of open and enclosed arm entries, and time spent on open arms were manually registered (13). Entry into an arm was defined as the point when the animal placed all four paws onto the arm. The percentage of open arm entries ( $100 \times \text{open}/\text{total entries}$ ) and the percentage of time spent in the open arms ( $100 \times \text{open}/(\text{open} + \text{enclosed})$ ) were calculated for each animal. Observations made from an adjacent corner produced significant ( $p < 0.001$ ,  $p < 0.05$ ) decreases of locomotion from its initial value during the period of the experiment (Table 1).

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Maximum suppression of locomotor activity was displayed at the dose of 400 mg/kg body weight, which was comparable to the reference drug diazepam.



**Figure: EPM test**

### **Statistical analysis:**

Experimental values are expressed as Mean  $\pm$  SEM. Independent Sample t-test was carried out for statistical comparison. Statistical significance was considered to be indicated by a P value  $<$  0.05 in all cases.

### **Extract preparation:**

The plant extract was prepared by blending and macerating 500g of the fresh leaves of *kalanchoe pinnata* with 100ml of distilled water and was kept at 40°C for 24 hours for extraction to take place. The resulting mixture was filtered. The concentration of the extract recovered from the filtrations was computed using the expression:

$$\text{Concentration} = (X - Y) / Z \text{ g/ml}$$

Where *X* = Weight of fresh leaves before blending

*Yg* = Weight of leaves after filtration

*Zml* = Volume of water after filtration

Fresh preparation was used for each experimental run.

### **Animals**

. Swiss albino mice of either sex, weighing 22-28 g, obtained from the Animal Resource Division, International Center for Diarrheal Disease and Research, Bangladesh (ICDDR), were used throughout the experiments. All animals were kept in standard environment condition, had free access to standard.

### **Chemicals :**

Glacial acetic acid was obtained from Sigma Chemicals, USA; diazepam was obtained from Square Pharmaceuticals Ltd., Bangladesh. All other chemicals were of analytical grade.

### **Animals**

. Swiss albino mice of either sex, weighing 22-28 g, obtained from the Animal Resource Division, International Center for Diarrheal Disease and Research, Bangladesh (ICDDR), were used throughout the experiments. All animals were kept in standard environment condition, had free access to standard food (ICDDR formulated) and water *ad libitum*. Prior to experiments, all the animals were acclimatized for one week at ambient temperature.



**Figure: Albino swiss mice**

### **Experimental design:**

The animals were divided into five groups. The first groups of animals were used for the behavioural changes and acute toxicity studies. The second groups were used for the exploratory behaviour. The third groups were used for the benzodiazepam sleeping time. The fourth group were used also for the muscle relaxant tests. (Grp IV). The group which is group five was used for the study of anticonvulsant activity of the aqueous extract. Each group of animals was subdivided into control and experimental groups, which was treated with the extract at sub-lethal doses of 50, 100, 200mg/kg bodyweight while the control received normal saline. The vehicle and the extracts were administered intra peritoneally before the experimentation



Neuropharmacological Evaluation of *k. pinnata***RESULT:****Acetic Acid induced writhing test****Table:** No. of writhing

	No. of writhing	% inhibition
POSITIVE	0.1±0.112***	99.70674
NEGATIVE	34.10±7.478	0
K. pinnata EA	14.80±4.186*	56.59824

The effect of the ethyl acetate extract of *k. pinnata* is comparable with standard and in some cases more effective than the standard.

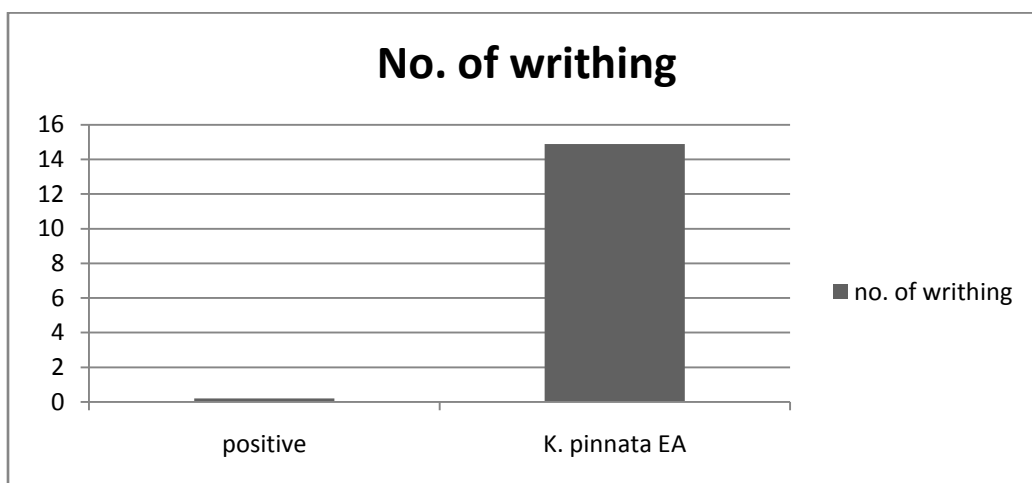


Figure:No. of writhing



Neuropharmacological Evaluation of *k. pinnata***Formalin induced biphasic pain****Table: licking time in early phase and late phase**

	Early phase	% inhibition	Late phase	% inhibition
positive	42.20 ± 2.219***	46.96429	25.4 ± 1.924**	24.05822
negative	110.4 ± 6.130	0	61.00 ± 4.472	0
<i>K. pinnata</i> EA	83.20 ± 2.71	20.35714	56.2 ± 3.156	38.61301

\*\*\*the mean difference is significant at the 0.001 level.

\*\*the mean difference is significant at the 0.01 level.

The effect of the ethyl acetate extract of *k. pinnata* is shown in the graph.

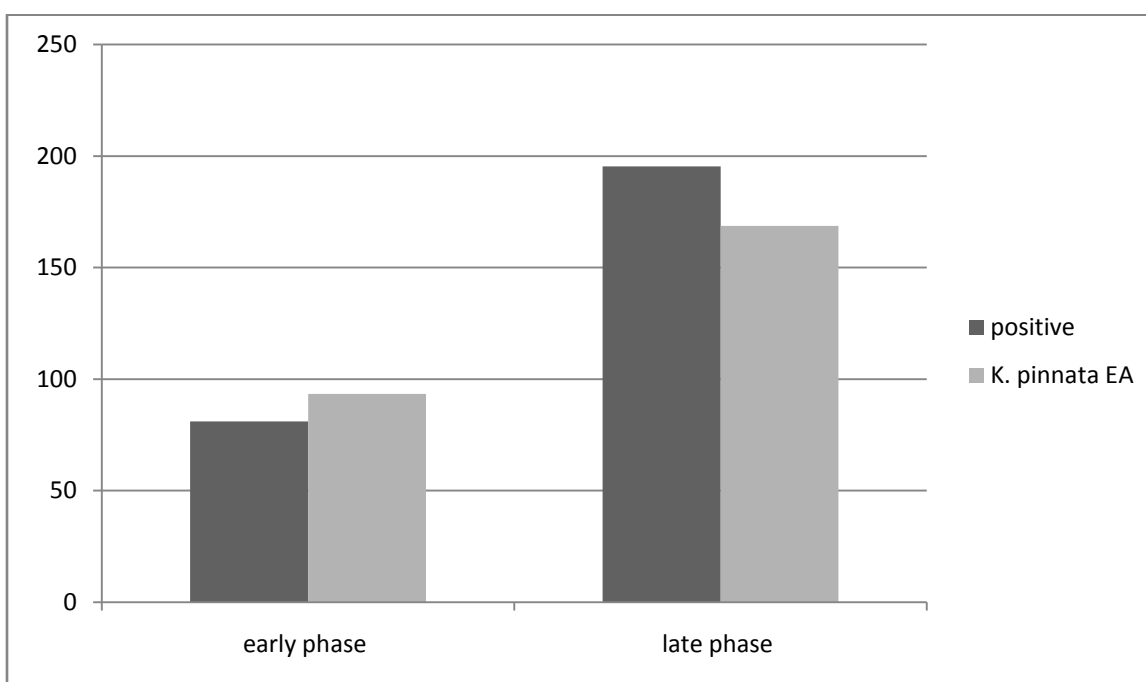


Figure: licking time of the early phase and late phase of the treated mice

Neuropharmacological Evaluation of *k. pinnata***HOLE CROSS test****Table : Number of Hole crossed by the mice**

Minute	Control	Diazepam	K.pinnata EA
0	18.60±0.570	12.80±3.090	11.40±1.643
30	9.20±2.329	6.60±1.151	5.60±1.440
60	5.40±0.671	4.40±1.483	2.60±1.351
90	5.40±0.570	4.60±0.837	1.40±0.837
120	4.60±0.570	3.80±1.917	0.80±0.224

The effect of the ethyl acetate extract of *k. pinnata* is comparable from the second interval with standard and in some cases more effective than the standard.

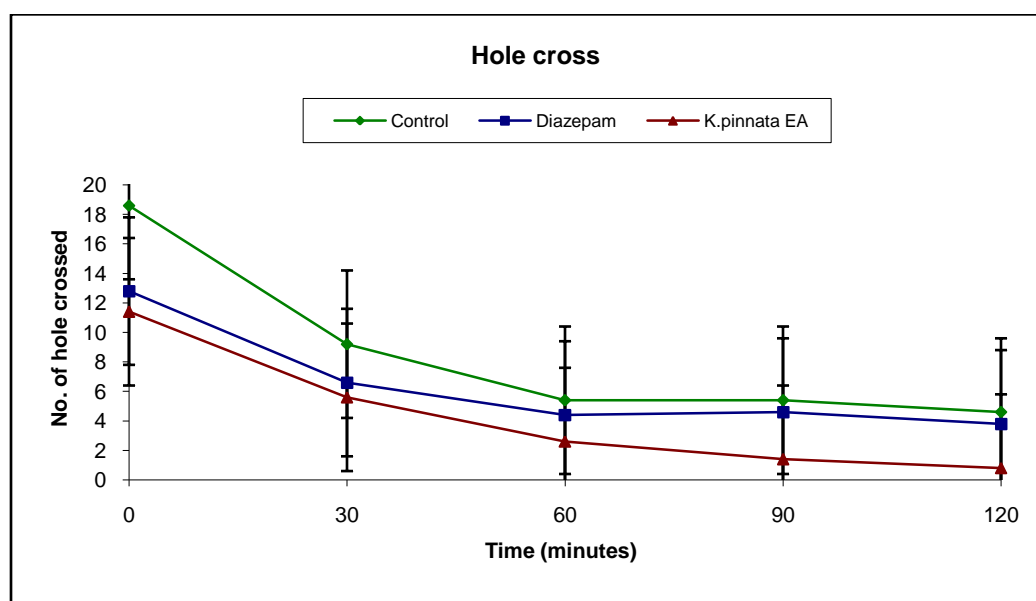


Figure: Number of Hole crossed by the mice

Neuropharmacological Evaluation of *k. pinnata***Open Field****Table: Number of square travelled by the mice**

Minute	Control	Diazepam	K.pinnata EA
0	121.20±0.7.652	110.40±5.943	85.00±5.099
30	60.20±3.435	56.20±7.012	31.80±2.104
60	40.60±4.207	36.60±3.616	25.00±1.969
90	29.40±2.864	25.80±2.302	20.00±0.791
120	28.80±4.866	24.80±8.863	22.40±2.797

The effect of the ethyl acetate extract of *k. pinnata* is comparable from the second interval with standard and in some cases more effective than the standard.

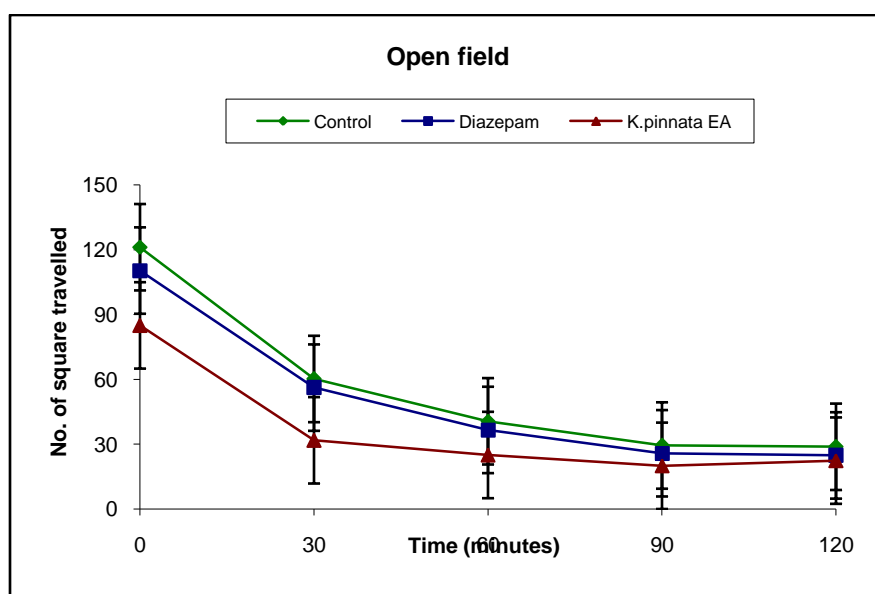


Figure: Number of square travelled by the mice

Neuropharmacological Evaluation of *k. pinnata***EPM****Table : % number of entry and time spent in the open arm by the mice**

	% no. of entry into the open arm	% time spent in the open arms
Positive	76.28±1.847**	79.39±5.749**
Negative	55.88±2.133	51.93±8.243
K. pinnata EA	17.00±3.687	2.71±0.346

\*\*the mean difference is significant at the 0.01 level.

The effect of the ethyl acetate extract of *k. pinnata* is comparable with standard and in some cases more effective than the standard.

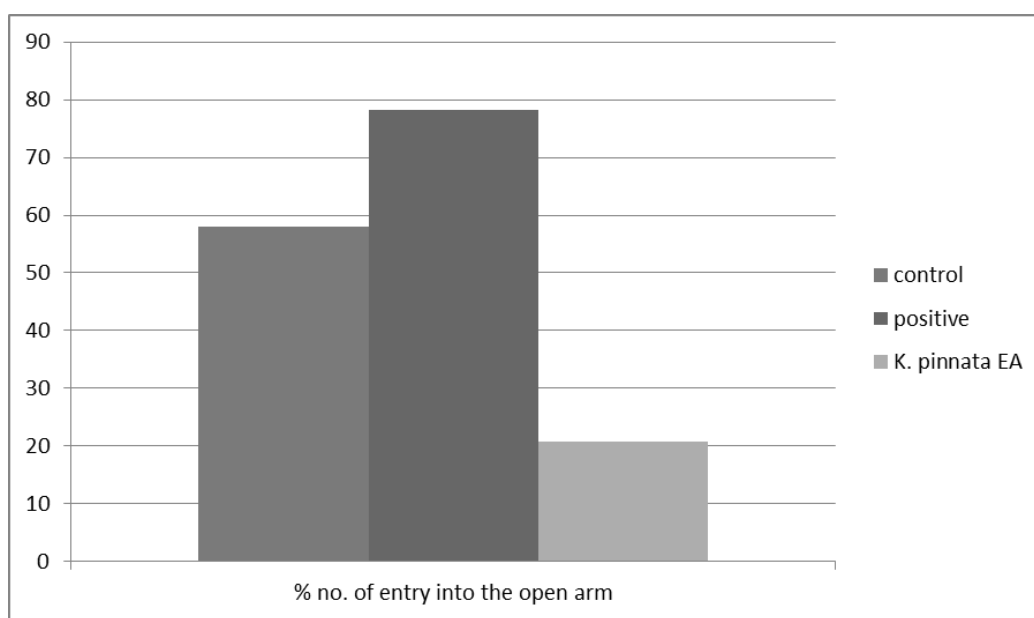


Figure: % number of entry into the open arm

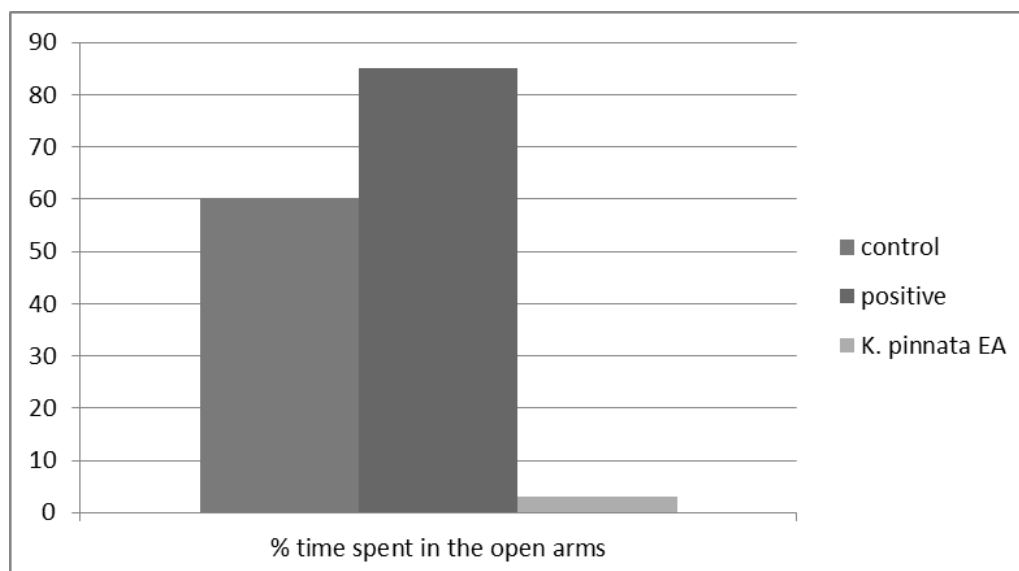


Figure: % time spent by the mice in the open arm

## Discussion

Acetic acid induced writhing is the main point for finding new peripherally active analgesic drugs (Hasan *et al.*, 2010). Pain sensation in writhing method is due to abdominal constrictions associated with irritation of peritoneal cavity by acetic acid. Prolonged acetic acid induced irritation lead to increase levels of prostaglandins (PGE<sub>2</sub> and PGF<sub>2</sub> $\alpha$ ) biosynthesis via cyclooxygenase (COX) and lipoxygenase products in peritoneal fluids followed by increase levels of free arachidonic acid secretion from tissue phospholipid. These increase levels of prostaglandins and lipoxygenase products enhance inflammatory pain by increasing capillary permeability in peritoneal cavity of abdomen (Zulfiker *et al.*, 2010; Zakaria *et al.*, 2008). The analgesic agents reduced the number of writhing preferably by inhibition of synthesis of prostaglandins and lipoxygenase products (Ferdous *et al.*, 2008). The crude extract showed significant analgesic action compared to the reference drug aspirin by reducing the number of acetic acid induced writhing in mice at dose of 400 mg/kg. Thus, the results indicate that the significant pain reduction of the plant extract might be due to the presence of peripherally active analgesic principles. Similarly in chimney, climbing and inclined screen tests, there was a significant loss of coordination and decrease muscle tone in animals treated intraperitoneally

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with *k. pinnata* aqueous extract in a dose dependent fashion (Table 2). The treatment with *k. pinnata* extract, is also able to cause a dose related delay of the onset in tonic and picrotoxin, even if it was unable to prevent convulsion, an inhibition of mortality was also observed with 400mg/kg. The result of the present study indicates that the crude extract of the *k. pinnata* leaf produced a significant alterations in general behaviour pattern, reduction in spontaneous mortality, potentiation of pentobarbitone-induced sleeping time in a dose dependent fashion. This is similar with the findings of Fujimori (1995) who proposed that the enhancement of barbital hypnosis is a good index of CNS depressant activity (Fujimori, 1995). The extract also produces a significant decrease in exploratory behaviour pattern as evident from the results of head-dip, climbing, and evasion tests. Furthermore the aqueous extract of *K. Pinnata* produces minor anticonvulsant effect by delaying seizure produced by strychnine and picrotoxin. It was however more efficacious against picrotoxin induced seizure where protection was observed in about one-quarter of mice, an effect which indicates that *B. pinnatum* aqueous extract might produce its central nervous system depressant action as consequence of its GABAergic and less importantly, glycinergic transmission, since picrotoxin is a selective GABAA receptor antagonist (Rang et al 1996) while strychnine antagonizes the inhibitory spinal cord and brainstem reflexes of glycine (Yemitan et al 2001).

## Conclusion

The acetic acid leaf extract of *kalanchoe pinnata* showed a significant analgesic activity in writhing test and a beneficial preventive effect in neuropharmacological test (open field test, maze test and hole tet). However, further study is needed in order to understand the precise mechanism. Studies with pure active compounds of the extract must be conducted for further pharmacological and toxicological characterization.



## References

1. Sharma, A., C. Shanker, L.K. Tyagi, M. Singh and C.V. Rao, Herbal medicine for market potential in India: An overview. Acad. J. Plant Sci., 2008, 1: 26-36.
2. Joseph, B., R.M. Priya P.A.M. Helen and S. Sujatha, Bio-active compounds in essential oil and its effects of antimicrobial, cytotoxic activity from the *Psidium guajava* (L.) Leaf. J. Adv. Biotechnol., 2010, 9: 10-14.
3. B. Joseph, S. Sridhar, Sankarganesh, Justinraj and Biby T. Edwin, Rare Medicinal Plant- *Kalanchoe Pinnata*. Research Journal of Microbiology, 2011, 6: 322-327.
4. <http://findmeasure.com/2009/03/25/kalanchoe-pinnata/>
5. [http://en.wikipedia.org/wiki/Kalanchoe\\_pinnata](http://en.wikipedia.org/wiki/Kalanchoe_pinnata)
6. <http://www.medicineatyourfeet.com/kalanchoepinnata.htm>.
7. P. Paranjpe. Indian Medicinal Plants forgotten Healers. Chaukhamba Sanskrit Pratisthan, 2Delhi,2005,194-195.
8. S. Jaiswal, and S. Sawhney. Correlation of epiphyllous bud differentiati on with foliar senescence in crassulacean succulent *Kalanchoe pinnata* as revealed by thidiazuron and ethrel application. J. of Plant Physiology. 2006, 163: 717-722.
9. P.B. Marriage, and D.G. Wilson. Analysis of Organic acids of *Bryophyllum pinnatum*. Can. J. Biochem. 1971, 49: 282-295.
10. K. Gaiind, and R. Gupta. Alkanes ,Alkanols, Triterpenes, and Sterols of *Kalanchoe Pinnata*. Phytochemistry. 1972, 11: 1500-1502.
11. K. Gaiind, and R. Gupta. Identification of waxes from leaves of *Kalachoe pinnata*. *Planta Medica*. 1974,23: 193-197.
12. R.A. McKenzie, F.P. Franke, and P.J. Dunster. The toxicity to Cattle and Bufadienolide content of six *Bryophyllum* species. Aust Vet J. 1987,64 (10): 298-301.
13. T. Yamagishi, M. Haruna, X.Z. Yan, J.J. Chang, and K.H. Lee. Antitumor agents, 110, *Bryophyllin B*, A Novel Potent cytotoxic Bufadienolide from *Bryophyllum Pinnatu.m* J. Nat. Prod. 1989, 52(5): 1071-1079.



Neuropharmacological Evaluation of *k. pinnata*

14. R.P. Rastogi, and B.N. Mehrotra. Compendium of Indian Medicinal Plants,1990-1994, 5: 141- 143.
15. Supratman, U., T. Fujita, K. Akiyaa, H. Hayashi and A. Murkami *et al.*, Anti-tumor promoting activity of bufadienolides from *Kalanchoe pinnata* and *K. daigremontiana* X *tubiflora*. *Biosci. Biotechnol. Biochem.*, 2001,65: 947-949.
16. Ram, P.R. and B.N. Mehrotra, 2004. Compendium of Indian Medicinal Plants, 2004. Vol. 5. ISBN: 81-85042-13-6.
17. U. Supratman, T. Fujita, K. Akiyama, and H. Hayashi. New insecticidal bufadienolide, Bryophyllin C from *Kalanchoe pinnata*. *Biosci Biotechnol Biochem.* 2000,64(6): 1310-1312.
18. D.A. Akinpelu. Antimicrobial activity of *Bryophyllum Pinnatum* leaves *Fitoterapia.* 2000, 71(2): 193-194.
19. D.E. Okwu, and C. Josiah. Evaluation of the chemical composition of two Nigerian medicinal plants. *African Journal of Biotechnology.* 2006,5(4): 357–361.
20. Toshihiro, K. WCMC, T. Toshitake, and M. Taro. Sterols of *Kalanchoe pinnata*. First report of the isolation of both C-24 epimers of 24-Alkyl-A25-sterol from higher plants. *Lipids.* 1991,26: 660 .
21. S. Hunt, I.L. Groff, and J. Holbrook. *Principles and Chemical Practice.* John Wiley and sons New York. 1980,459-462.
22. K.C. Ofokansi, C.O. Esimone, and C.R. Anele. *Plant Products Research Journal.* 2005,9: 23-27. Pattewar /2012 8
28. H. Fujimori. Potentiation of barbital hypnosis as an evaluation method for CNS depressant. *Psychopharmacology.* 1995,7: 374-377.
29. S. Pal, T. Sen, and A.K.N. Chaudhuri. Neuropsychopharmacological profile of the methanolic fraction of *Bryophyllum Pinnatum* leaf extract. *Journal of Pharmacy and Pharmacology.* 1999,51(3): 313-318.
30. H.M. Salahdeen, and O.K. Yemitan. Neuropharmacological Effects of Aqueous Leaf Extract of *Bryophyllum Pinnatum* in Mice. *African Journal of Biomedical Research.* 2006,9: 101-107.
31. B. Joseph, S. Sridhar, Sankarganesh , Justinraj and Biby T. Edwin,Rare Medicinal Plant- *Kalanchoe Pinnata.* *Research Journal of Microbiology,* 2011,6: 322-327.
32. C.Z. Nassis, E.M. Haebisch, and A.M. Giesbrecht. Antihistamine activity of *Bryophyllum Calycinum.* *Braz J. Med Bio. Res.* 1992,25(9):929-936.

Neuropharmacological Evaluation of *k. pinnata*

33. J.K. Adesanwo, Y. Raji, S.B. Olaleye, S.A. Onasanwo, O.O. Fadare, O.O. Ige, and O. O. Odusanya. Antiulcer Activity of Methanolic Extract of Bryophyllum pinnatum in Rats, Journal of Biological Sciences. 409-412.
34. E.E. Obaseiki-Ebor. Preliminary report on the invitro antibacterial activity of Bryophyllum pinnatum leaf juice. Afr J Med Med Sci. 1985,14(3-4):199-202.
35. J.A.O. Ojewole. Antinociceptive, anti-inflammatory and antidiabetic effects of Bryophyllum pinnatum (Crassulaceae) leaf aqueous extract. Journal of Ethno pharmacology. 2005,99: 13-19.
36. B. Rossi-Bergmann, S.S. Costa, M.B.S. Borges, S.A. da Silva, G.R. Noleto, M.L.M. Souza, and V.L.G. Moraes. Immunosuppressive effect of the aqueous extract of Kalanchoe Pinnata in mice. Phytothera. Res. 1994, 8: 399-402.
37. P. Siddharta, and A.K.N. Chaudhuri. Further studies on the Anti-inflammatory profile of the Methanolic Fraction of the fresh leaf extract of Bryophyllum Pinnatum. Fitoterapia.1992,63(5): 451-459 .
38. Nayak, B.S., J.R. Marshall and G. Isitor,. Wound healing potential of ethanolic extract of Kalanchoe pinnata Lam. leaf--a preliminary study. Indian J. Exp. Biol., 2010,48: 572-576.
39. B. Gwehenberger, L. Rist, R. Huch, and U. von Mandach. Effect of Bryophyllum pinnatum versus fenoterol on uterine contractility. Eur J Obstet Gynecol Reprod Bio1. 2004,113(2):164-171.
40. G.P. Reppas. Bryophyllum Pinnatum poisoning of Cattle. Aust Vet J. 1995, 72(11): 425-427
41. D.A. Alabi, I.A. Oyero, Jimoh and N.A. Amusa. Fungitoxic and Phytotoxic Effect of Vernonia amygdalina (L), Bryophyllum pinnatum Kurz Ocimum gratissimum (Closium) L. and Eucalyptna globules (Caliptos) Labill Water Extracts on Cowpea and Cowpea Seedling Pathogens in Ago Iwoye, South Western Nigeria. World Journal of Agricultural Sciences. 2005,1(1): 70-75.
42. Willcox, M.L. and G. Bodeker, Traditional herbal medicines for malaria. BMJ, 2004. 329: 1156-1159.